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# MOLECULAR DESIGN OF NOVEL PGI2 AGONISTS WITHOUT PG SKELETON. I

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**Abstract.** Syntheses of novel prostaglandin (PG) mimetics without PG skeleton are described, and the structure-activity relationships are discussed. Highly potent compounds could be obtained when the PG analog 3 was modified by removing and reconstructing the cyclopentane ring, and changing the allylic alcohol in the natural PGs.

Since the discovery of prostacyclin  $(PGI_2, 1)$ ,  $^1$  many efforts have been devoted to the search for the chemically stable, biologically potent and tissue specific agonists for potential medical utility.  $^2$  PGI<sub>2</sub> and PGE<sub>1</sub> (2) have a different structure, however, they bind to the same platelet PGI<sub>2</sub> receptors and show qualitatively the same biological activities.  $^3$  For these biological activities of PGs, the carboxylic acid, the cyclopentane ring and the allylic alcohol moieties have been demonstrated to be essential.  $^4$  Accordingly, approaches to their specific agonists were so far limited to the structural modifications of the  $\alpha$  and  $\omega$ -chains in PGs. Recently, an Edinburgh University group and the Bristol-Myers Squibb group, respectively, proposed EP-157<sup>5</sup> and BMY 42393<sup>6</sup> as the PGI<sub>2</sub> agonists without PG skeleton.

We were greatly concerned with the role of the cyclopentane ring. The cyclopentane ring involving the 11-hydroxyl group, 9-oxygenated function, and 8R and 12S configurations of PGI<sub>2</sub> and PGE<sub>1</sub> have been considered to bind to the receptors. At first, we assumed that the cyclopentane ring controlled the geometrical relationship between the carboxylic acid and the allylic alcohol functions in PGs. If the new structures are able to consist of the same spatial arrangement as that of the active conformer in PGs, it would be reasonable to assume that they should show the biological activities comparable to those of PGs.

We began a research program with the goal of identifying a new  $PGI_2$  agonist without these cyclopentane ring and allylic alcohol parts. We selected the  $PGE_1$  analog  $3^7$  for the study of a non PG-like structure. Our

strategy as shown in Figure 1 is to design molecules in which the cyclopentane ring part in PG is removed to construct a new skeleton (4a-c), and in which the allylic alcohol part in PG is modified to show the PGI<sub>2</sub>-like activities (9b).

# Chemistry

The compounds were synthesized by the routes shown in Schemes 1, 2 and 3. The synthesis of **4a** and **9a** is depicted in Scheme 1. Commercially available 3-hydroxyphenyl ethanol **10** was converted in six steps to ester aldehyde **11**. Wadsworth-Emmons reaction<sup>8</sup> of **11** produced enone, which was converted to **4a** by the reduction with sodium borohydride in the presence of catalytic amount of acetic acid followed by saponification. Oximization of **11** with benzhydryloxy amine<sup>9</sup> and saponification gave **9a**.

(1) SO<sub>3</sub>•pyridine, Et<sub>3</sub>N, DMSO; (2) Ph<sub>3</sub>PCHCOOMe, CHCl<sub>3</sub>; (3) H<sub>2</sub>, Pd-C, AcOEt; (4) LiAlH<sub>4</sub>, THF; (5) K<sub>2</sub>CO<sub>3</sub>, BrCH<sub>2</sub>COOMe, MeCN; (6) Swern Ox.; (7) LiCl, *i*-Pr<sub>2</sub>NEt, (MeO)<sub>2</sub>POCH<sub>2</sub>CO(CH<sub>2</sub>)<sub>4</sub>CH<sub>3</sub>, MeCN; (8) NaBH<sub>4</sub>, cat.AcOH, MeOH, -20°C; (9) aq.NaOH, MeOH; (10) H<sub>2</sub>NOCHPh<sub>2</sub>, EtOH.

2-Substituted-tetrahydronaphthalene-5-oxyacetic acids 4b-9b were prepared as shown in Scheme 2. Compound 13 was easily accessible starting from 5-methoxy-2-tetralone 12<sup>10</sup> by Reformatsky reaction 11 followed by dehydration and hydrogenation. Key intermediate 14 was obtained by the following series of

reactions: (i) reduction of 13 with lithium aluminum hydride to afford alcohol; (ii) acylation; (iii) demethylation; (iv) saponification; (v) O-alkylation; and (vi) oxidation. Compounds 4b, 5b, and 9b were easily prepared by the same preceding procedure. Wittig reactions of 14 with two types of the ylide furnished stereoselectively ciscompounds which were contaminated with a trace of trans-isomers. This trans-isomers were readily isomerized by the reaction of light. Saponification gave 6b, 7b, and 8b.

#### Scheme 2

(1) Zn, BrCH<sub>2</sub>COOEt, C<sub>6</sub>H<sub>6</sub>; (2) POCl<sub>3</sub>, pyridine; (3) H<sub>2</sub>, Pd-C, AcOEt; (4) LiAlH<sub>4</sub>, THF; (5) Ac<sub>2</sub>O, pyridine; (6) BBr<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>; (7) aq.NaOH, MeOH; (8) K<sub>2</sub>CO<sub>3</sub>, BrCH<sub>2</sub>COOMe, MeCN; (9) Swern Ox.; (10) (MeO)<sub>2</sub>POCH<sub>2</sub>CO(CH<sub>2</sub>)<sub>4</sub>CH<sub>3</sub>, LiCl, *i*-Pr<sub>2</sub>NEt, MeCN; (11) NaBH<sub>4</sub>, cat.AcOH, MeOH, -20°C; (12) aq.NaOH, MeOH; (13) (MeO)<sub>2</sub>POCH<sub>2</sub>COCH<sub>2</sub>Ph, LiCl, *i*-Pr<sub>2</sub>NEt, MeCN; (14) Ph<sub>3</sub>P(CH<sub>2</sub>)<sub>3</sub>Ph, *n*-BuLi, THF; (15) hv, (PhS)<sub>2</sub>; (16) Ph<sub>3</sub>P(CH<sub>2</sub>)<sub>2</sub>CHPh<sub>2</sub>, *n*-BuLi, THF; (17) H<sub>2</sub>NOCHPh<sub>2</sub>, EtOH.

Scheme 3 illustrates the preparation of 1-substituted-tetrahydronaphthalene-5-oxyacetic acids 4c and 9c. Conversion of commercially available 5-methoxy-1-tetralone 15 into 16 was achieved by (i) treatment with trimethylsilyl nitrile to give cyanohydrin; (ii) dehydration with phosphorous oxychloride to  $\alpha,\beta$ -unsaturated nitrile; (iii) diisobutylaluminum hydride reduction; (iv) Wittig reaction; and (v) hydrogenation. 16 was transformed to ester-aldehyde 17 in six steps. Compounds 4c and 9c were easily prepared by the same preceding procedure.

## Biological Results and Discussion

In order to assess the affinities of the target compounds for platelet PGI<sub>2</sub> receptors, radioligand binding assays were performed in human platelets using [<sup>3</sup>H]-iloprost.<sup>3</sup>

Table 1 shows the affinities of simple allylic alcohol compounds. Normal alkyl compound 4a, in which the ring was removed from the structure 3, bound weakly to the PGI<sub>2</sub> receptors. This weakness can be

(1) Me<sub>3</sub>SiCN; (2) POCl<sub>3</sub>, pyridine; (3) DIBAL, PhCH<sub>3</sub>, and then H<sub>2</sub>SO<sub>4</sub>; (4) Ph<sub>3</sub>PCHCOOMe, CHCl<sub>3</sub>; (5) H<sub>2</sub>, Pd-C, AcOEt; (6) LiAlH<sub>4</sub>, THF; (7) Ac<sub>2</sub>O, pyridine; (8) BBr<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>: (9) aq.NaOH, MeOH; (10) K<sub>2</sub>CO<sub>3</sub>, BrCH<sub>2</sub>COOMe, MeCN; (11) Swem Ox.; (12) LiCl, *i*-Pr<sub>2</sub>NEt, (MeO)<sub>2</sub>POCH<sub>2</sub>CO(CH<sub>2</sub>)<sub>4</sub>CH<sub>3</sub>, MeCN; (13) NaBH<sub>4</sub>, cat.AcOH, MeOH, -20°C; (14) aq.NaOH, MeOH; (15) H<sub>2</sub>NOCHPh<sub>2</sub>, EtOH.

attributed to the high flexibility of 4a, which presumably prevented it from binding. To circumvent this, the introduction of an additional ring to diminish the flexibility of 4a gave the 2,5- and 1,5-disubstituted tetrahydro naphthalene derivatives 4b and 4c with IC<sub>50</sub>'s of 7.8  $\mu$ M and 9.6  $\mu$ M, respectively.

These results gave an argument that the cyclopentane ring part would control the relationship between the carboxylic acid and the allylic alcohol moieties in PG, and the relative relationship between the carboxylic acid and the allylic alcohol moieties in **4b** and **4c** would be similar to that in PGI<sub>2</sub> and PGE<sub>1</sub>.

**Table 1.** The Effect without Cyclopentane Ring on the Displacement of [<sup>3</sup>H]-lloprost from Human Platelet PGI<sub>2</sub> Receptor

No.		IC <sub>50</sub> (μM)
<b>4</b> a	ОСООН	23
4b	ОСООН	7.8
4c	ОСООН	9.6

We next turned our attention to the allylic alcohol moiety as shown in Table 2. In order to examine whether 4b would show PG-like behavior, 4b was first modified in a classical way as in PGs. A phenyl ring

was introduced to the  $\omega$ -chain to afford 5. As in the case of PG, this modification caused a little change in the receptor binding affinity. The IC50 value of compound 5 was 4.9  $\mu$ M. However, the removal of the hydroxyl group in 5 gave the interesting compound 6 with the IC50 value of 5.3  $\mu$ M. As opposed to prostaglandins, it is still active without the allylic alcohol. Further modifications of compound 6 led to compound 7 with the IC50 value of 3.1  $\mu$ M. At this stage, its *cis* isomer 8 showed the same binding potency. Therefore, we considered that the binding affinity should be dependent on the geometrical relationship between the carboxylic acid and the terminal phenyl groups. This prompted us to search for other functional groups with more potent binding affinity than the double bond. When the double bond in compound 8 was replaced by an oxime moiety, the potency of resulting compound 9b remarkably increased to IC50 of 0.65  $\mu$ M!

**Table 2.** The Effect of Variation of the Allylic Alcohol Part on the Displacement of [<sup>3</sup>H]-lloprost from Human Platelet PGI<sub>2</sub> Receptor

No.	X	IC <sub>50</sub> (μΜ)
4b	TE OH	7.8
5b	רק ׄPh OH	4.9
6b	Ph Ph	5.3
7b	Ph	3.1
8b	Ph Ph	3.0
9b	مرچ <sup>N.</sup> O Ph	0.65

The skeletal changes of the oxime analogs gave IC<sub>50</sub> values of 3.9  $\mu$ M for 9a and 0.60  $\mu$ M for compound 9c in Table 3. It is interesting to note the similarity in the binding tendency of these oxime ethers, as compared to those of compounds possessing the allylic alcohol, as we mentioned before.

The IC50 values of inhibition of 4  $\mu$ M ADP-induced human platelets aggregation were 1.1  $\mu$ M for 9b, and 2.1  $\mu$ M for 9c.

Notably, the agents detailed herein lack the cyclopentane ring and the allylic alcohol in PGs have been which have been long considered essential for the biological activity. Thus, though a reassessment of the role of each functionality included in the PG skeleton, we succeeded in the design of the new PGI<sub>2</sub> agonists.

No.		IC <sub>50</sub> (μM)
9a	N. o Ph	3.9
9b	O COOH Ph	0.65
9c	O_COOH Ph	0.60
lioprost		0.027

**Table 3.** The Effect of Skeletal Change on the Displacement of [<sup>3</sup>H]-lloprost from Human Platelet PGl<sub>2</sub> Receptor

## References and Notes

- (a) Moncada, S.; Gryglewski, R. J.; Bunting, S.; Vane, J. R. Nature, 1976, 263, 663. (b) Gryglewski, R. J.; Bunting, S.; Moncada, S.; Flower, R. J.; Vane, J. R. Prostaglandins, 1976, 12, 685. (c) Johnson, R. A.; Morton, D. R.; Kinner, J. H.; Gorman, R. R.; McGuire, J. C.; Sun, F. F.; Whittaker, N.; Bunting, S.; Salmon, J.; Moncada, S.; Vane, J. R. Prostaglandins, 1976, 12, 915.
- 2. Muchowski, J. M. Handbook of Eicosanoids: Prostaglandins and Related Lipids; Willis, A. L., Ed.: 1987; Vol. 1, Part B, pp. 42-81.
- 3. Schafer, A. I.; Cooper, B.; O'Hara, D.; Handin, R. I. J. Biol. Chem., 1979, 254, 2914.
- 4. Nickolson, R. C.; Town, M. H.; Vorbruggen, H. Med. Res. Rev., 1985, 5, 1.
- Muir G., Jones R.L., Will S.G., Winwick T., Peesapatil V., Wilson N.H., Griffiths N., Nicholson W.V., Taylor P., Sawyer L., Blake A.J., Eur. J. Med. Chem., 1993, 28, 609.
- 6. Meanwell, N. A.; Rosenfild, M. J.; Trehan, A. K.; Romine, J. L.; Wright, J. J. K.; Brassard, C. L.; Buchanan, J. O.; Federici, M. E.; Fleming J. S.; Gamberdella, M.; Zavoico, G. B.; Seiler, S. M. J. Med. Chem., 1992, 35, 3498.
- 7. Nelson, N. A.; Jackson, R. W.; Au, A. T.; Wynalda, D. J.; Nishizawa, E. E. *Prostaglandins*, 1975, 10, 795.
- 8. Blanchette, M. A.; Choy, W.; Davis, J. T.; Essenfeld, A. P.; Masamune, S.; Roush, W. R.; Sakai, T. *Tetrahedron Lett.*, **1984**, 25, 2183.
- 9. Grochowski, E.; Jurczak, J. Snthesis, 1976, , 682.
- 10. Soffer, M. D.; Bellis, M. P.; Gellerson, H. E.; Stewart, R. A. Org. Synth., 1952, 32, 97.
- 11. Han, B. H.; Boudjouck, P. J. Org. Chem., 1982, 47, 5030.